TERRECO Workshop, Bayreuth, April 11 - 14, 2010

Flux Regulation, N Balances and Production in Agroecosystems of Haean Catchment

Integrated experiment – May-August 2010

Sina Berger, Laboratory for Isotope Biogeochemistry Janine Kettering, Dept. of Agroecosystem Research Bora Lee, Dept. of Plant Ecology Steve Linder, Dept. of Plant Ecology Emily Martin, Dept. of Animal Ecology



Objective

Understand ecosystem fluxes and measure their impact on:

- 1) Environmental sustainability
- 2) Ecosystem service provision

Objective

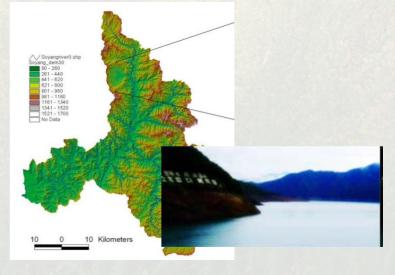
Understand ecosystem fluxes and measure their impact on:

- 1) Environmental sustainability
- 2) Ecosystem service provision

What environmental sustainability?

Atmospheric / Soil-plant **CO**₂ exchanges

N deposition, leaching and emissions



Objective

Understand ecosystem fluxes and measure their impact on:

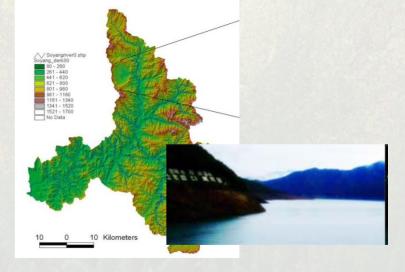
- 1) Environmental sustainability
- 2) Ecosystem service provision

What environmental sustainability?

→ Estimate and model the contribution of agricultural fields to the export of nutrients in the Soyang Lake reservoir

Atmospheric / Soil-plant **CO**₂ exchanges

N deposition, leaching and emissions



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Understand ecosystem fluxes and measure their impact on:

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What ecosystem services?

Agricultural **Production** Biological **Pest control**



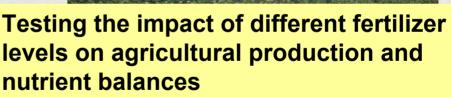
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Understand ecosystem fluxes and measure their impact on:

- 1) Environmental sustainability
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What ecosystem services?

→ Estimate optimal gains in ecosystem services of agricultural production, vs. limited impacts on water quality and nutrient balances Agricultural **Production** Biological **Pest control**







Main assumption

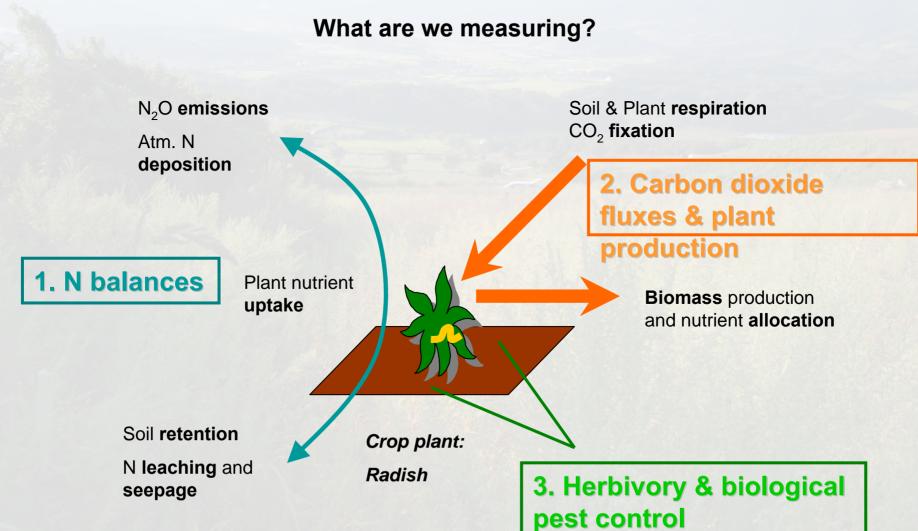
Ecosystem processes & fluxes both impact functioning and *interact with each* other

- Separate measurements of each process cannot account for such interactions
- ➔ In order to fully apprehend the set of parameters that influence production and sustainability, an <u>interdisciplinary approach</u> is necessary

Integrated approach to the measurement of ecosystem processes

Use of an identical field setup with coordinated measurements by multiple disciplines







- I. Experimental setup
- II. Nutrient cycling: N fluxes and N balances J. Kettering, S. Berger
- III. CO₂ fluxes and plant production S. Lindner, B. Lee
- IV. Herbivory and pest control E. Martin

I. Experimental setup

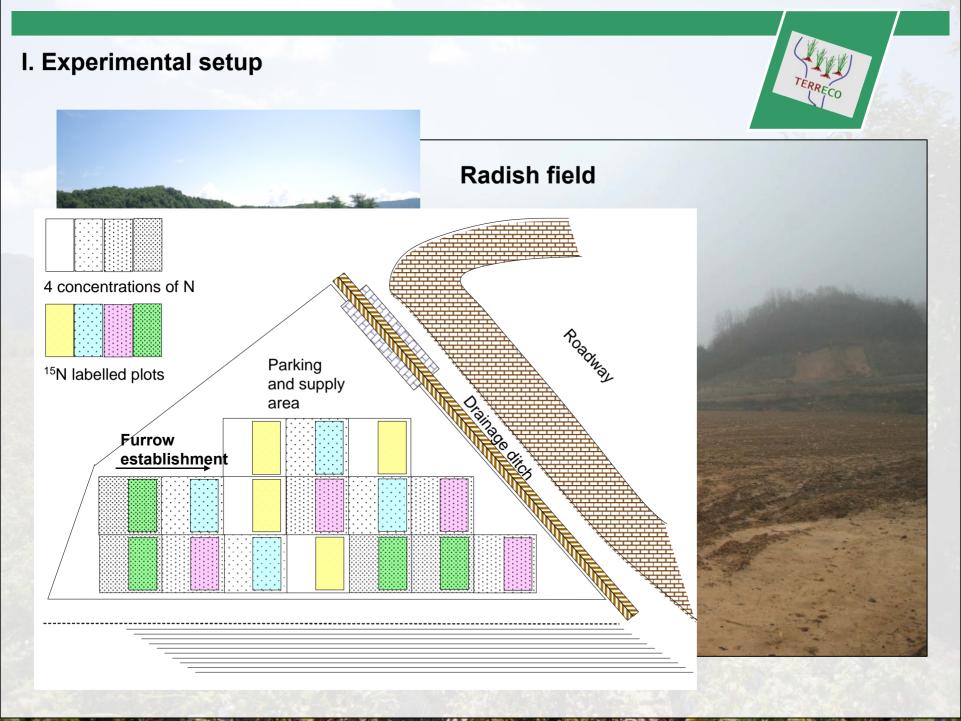


I. Experimental setup





Radish field No slope X ha



I. Experimental setup

1st fertilizer application

Ploughing

May

Disking

Ridges

Black

cover

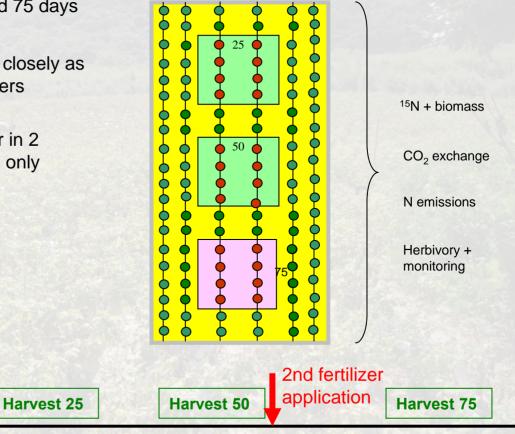
- 16 plots = 4 * 4 fertilizer levels
 → 50 150 250 350 kg N/ha
- Harvest of subplots after 25, 50 and 75 days
- Fertilizer application: reproduce as closely as possible the practices of local farmers
- Liquid or granulate *mineral* fertilizer in 2 applications (1st: everywhere, 2nd: only interrow), done manually

Planting (seeds)

June

Recommendation of Korean Agricultural Center: up to 400 kg N/ha

Usual amount in Germany: 50-150 kg N/ha



August

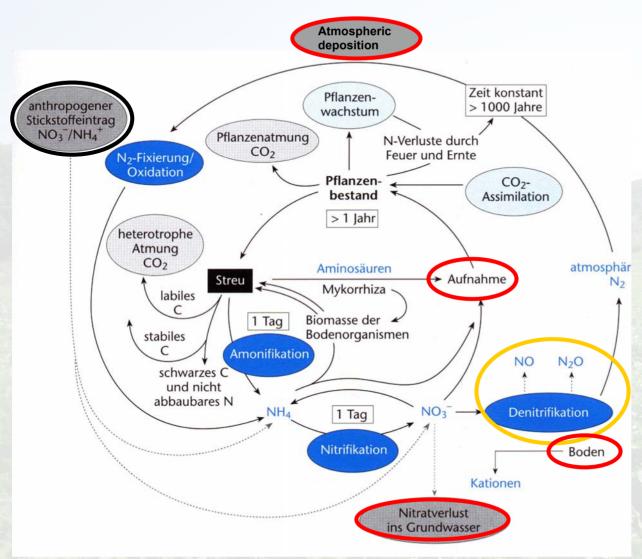
July



II. NUTRIENT CYCLING: N fluxes and N balances J. Kettering, S. Berger



II. NUTRIENT CYCLING – N fluxes and N balances



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<u>Input:</u> -Synthetic Fertilizer -Atmospheric deposition

Output: -Emissions -Seepage -Crop uptake -Retention in soil

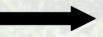
Janine
 Sina
 collective

Understand cycling of Nitrogen in agroecosystems

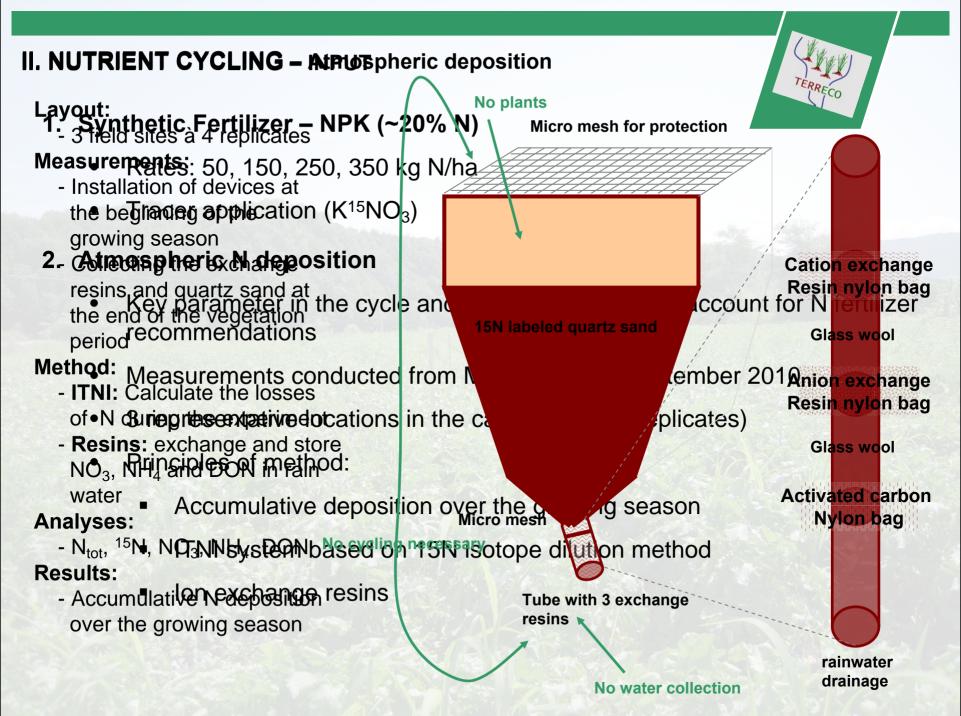


-How is N uptake by plants and N allocation affected by different N application rates? Does N use efficiency change?

- -How differs the biomass in response to different N application rates?
- -Which ones are the main N loss pathways and how do they differ in response to different N application rates?
- -How are the nitrous oxide emissions and leaching to the groundwater affected by different levels of N applied?
- -When do the biggest losses occur?
- -How soon after fertilizer application are huge amounts of the fertilizer already degassed as nitrous oxide?
- -How are nitrous oxide emissions influenced by the black cover?



Understand shifting of the cycle in response to different fertilizer levels



II. NUTRIENT CYCLING - OUTPUT

- 1. Retention in soil
- 2. Crop uptake
- 3. Seepage
- 4. Emissions

Frequency of measurements:

Seed



Soil sampling Atmospheric deposition



anvoet

Seepage

-3 harvest times

Fertilization

- -Soil (15N abundance)
- -Biomass
- -N uptake by plants (15N)
- -Seepage (15N)

-additionally

- -Nmin
- -Atmospheric deposition
- -Seepage after heavy rain events (15N)

Soil sampling for soil nutrient status

Day 50 2nd Fertilization

 Retention rate: percentage of applied 15N fertilizer recovered in the top 60 cm of the soil profile

Harvest

Harvest for N uptake

25

- Recovery rate: percentage of applied 15N fertilizer taken up by the

aboveground plant parts

- Root to shoot ratio with growth at differing fertilizer levels
- N use efficiency
- Biomass at different fertilizer levels

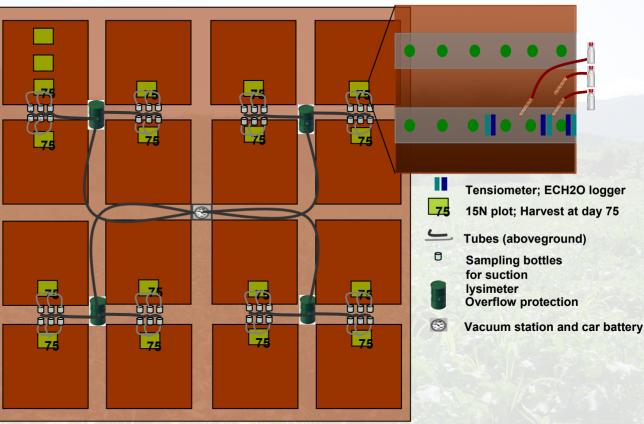
II. NUTRIENT CYCLING – SEEPAGE

Method:

-Suction lysimeter controlled by Tensiometers and FDR sensors for recording soil water content

-Disadvantage: don't capture preferential flow, non-continuous sampling, undefined soil volume

- -Advantage: easy to install and measure
- -3 suction lysimeter and tensiometer in each 75 days plot
- -Different depths and locations
- -4 sampling times plus after heavy rain events



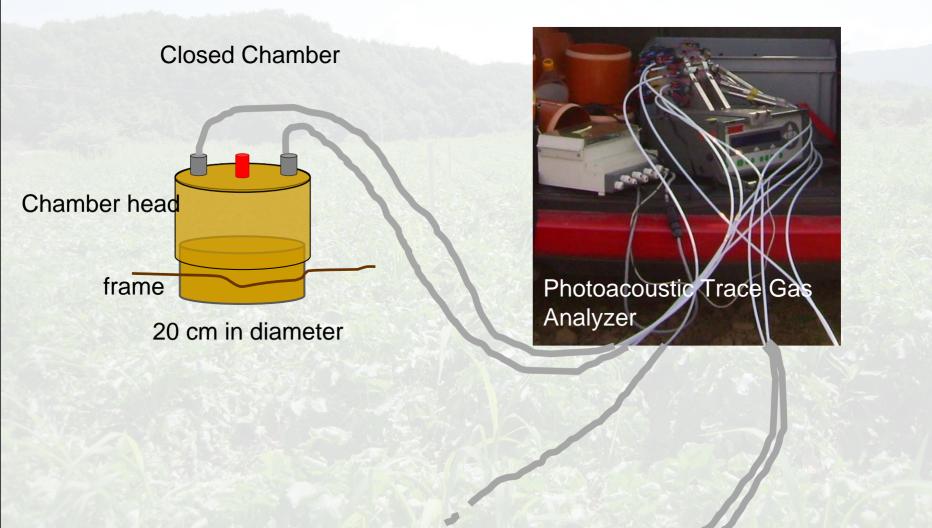
Results:

- -Modeled seepage over the course of a season
- -N loss in seepage water
- -Soil water content

II. NUTRIENT CYCLING – N₂O emission measurements



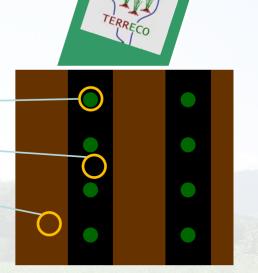
Method: Closed Chamber in conjunction with Infrared Photoacoustic Trace Gas Analyzer



II. NUTRIENT CYCLING – N₂O emission measurements

Design: On each of the plots three chambers will be installed:

- on a radish plant
- on black cover
- on path between rows



It will take one day to measure the N₂O emissions of all plots.

Measurement frequency:

Every day.....Reducing measurement days.....Every day...Reducing..Every day.... (measurements once a week)



Results:

- Nitrous oxide fluxes beetween soil and atmosphere
- Cumulative nitrous oxide emissions

III. Carbon dioxide fluxes and plant production S. Lindner, B. Lee

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Main objectives



1. Understand the impact of different input levels on CO₂ fluxes and plant production

\rightarrow In terms of LA, biomass, C/N content, carboxylation capacity & light use efficiency

2. Up scaling of CO₂ fluxes up to landscape level TERRECO-02: Spatial assessment of atmosphere-ecosystem exchanges via micrometeorological measurements, footprint modelling and mesoscale simulations Peng Zhao, Johannes Lüers, Thomas Foken, Chong Bum Lee

3. Validation of the Pixgro model

TERRECO-15: Comparisons of net ecosystem CO₂ exchange, carbon gain, growth and water use efficiency of agricultural crops in small catchments in Korea Bora Lee, John Tenhunen, Sinkyu Kang



Gas exchange data from chamber measurement and eddy system

Crop biomass data from crop sampling Land cover map from field survey and remote sensing

Meteorological data from AWS PIXGRO for Haean cahtchemnt Crop management data from fertilizer measurement

Introduction:

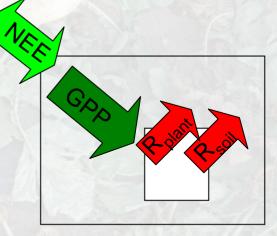


Dark chamber:

Soil respiration $R_{soil} = CO_2$ release from the bare soil **Ecosystem respiration** $R_{eco} = CO_2$ release from the soil (R_{soil}) + plant (R_{plant})

Light chamber:

Net ecosystem exchange NEE = GPP + Reco



Gross primary production (GPP): rate at which an ecosystem's producers capture and store a given amount of chemical energy as biomass in a given length of time.

Methods: Portable closed chamber system



Figure 1: Applied light and dark gas exchange chambers for measuring the NEE and R_{eco}



Figure 2: Installed soil frames $(38 \times 38 \text{ cm}^2)$ as a base for the gas exchange chambers

- Daily courses
- At least 5 times/ growing season and fertilizer level
- Intensified measurements on the Radish field with different fertilizer treatments
- NEE, Reco, Rsoil
- Microclimate
- Biomass leaves/ stem/ roots
- C/N content

- Detailed information of plant reaction to environmental factors in small scale (1-2 plants enclosed)

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Methods: CO2/H20 porometer CQP-130, Fa. WALZ, Effeltrich, Germany



- Measuring leaf gas exchange (photosynthesis or respiration of the leaf can be measured)

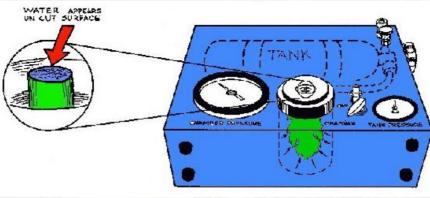
- In relation to microclimate

Methods: Pressure Chamber & Ech2o logger





- Plant water relations will be accessed using the Scholander pressure chamber



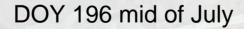


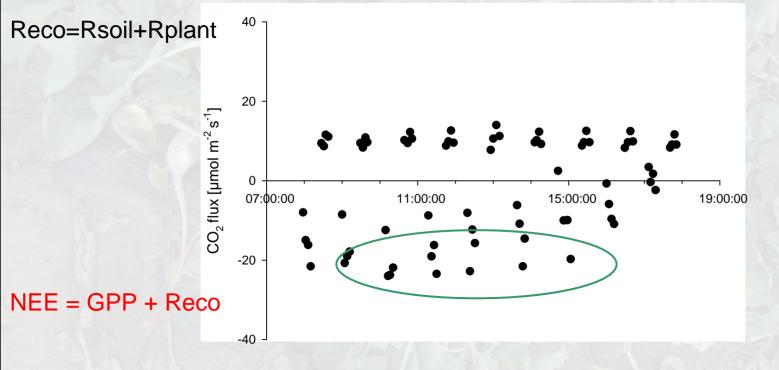
 Soil moisture content and soil temperature
 Automatic Weather Station for continuous recording of climate parameters (air temperature, relative humidity, solar radiation, wind speed and direction, rainfall)



Results from 2009:

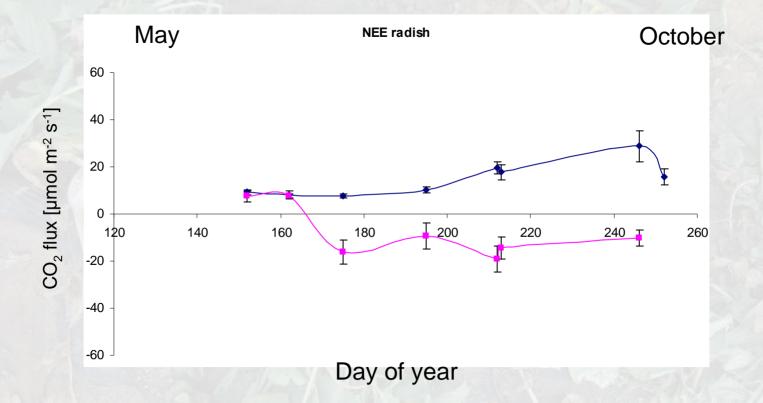






Daily course of NEE from a radish field

Results:

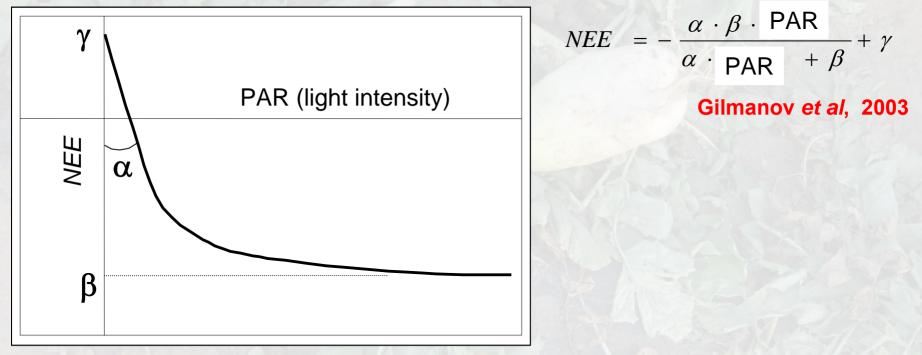


Seasonal course of CO₂ fluxes from radish

Hyperbolic light response model (Michaelis-Menten type model)



- Used Michaelis - Menten / rectangular hyperbola model to estimate model parameters for ecosystem/ leaf level gas exchange



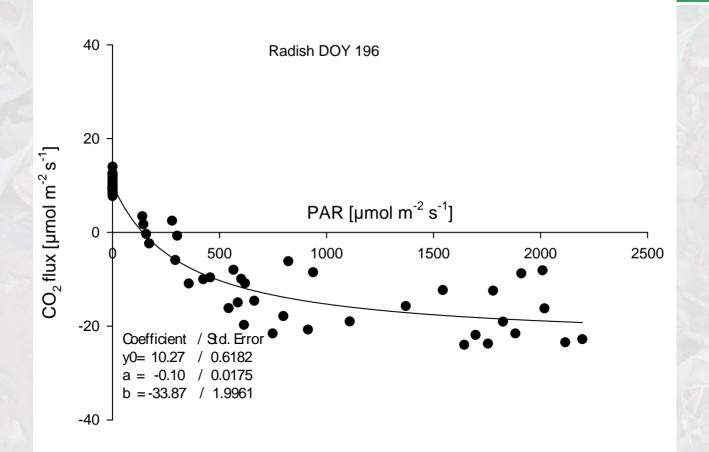
Physiological parameters:

Q is the initial slope of the light response curve and an approximation of the canopy light utilization efficiency

 $\boldsymbol{\beta}$ is the maximum NEE of the canopy

Y is an estimate of the average ecosystem respiration (Reco) occurring during the observation period

Results:



- Estimated parameters to describe gas exchange capacity of radish

IV. Herbivory and pest control E. Martin



IV. Herbivory and pest control



Introduction

Agricultural production is regulated by **bottom-up** and **top-down** processes

Abiotic constraints Resource limitation IV. Herbivory and pest control



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Agricultural production is regulated by **bottom-up** and **top-down** processes

Abiotic constraints Resource limitation

Availability of nutrients

/ Level of N fertilizer

IV. Herbivory and pest control

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Herbivore pressure

Natural predator compensation

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Herbivore pressure Natural predator compensation



Insects and birds act as natural enemies of pests by regulating insect herbivore populations

Pest mortality is induced by

- consumption (natural predators)
- parasitism (parasitoids wasps)



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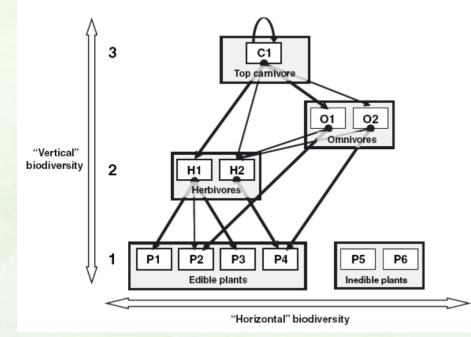
Natural predator compensation

Biological pest control

Multiple trophic level interactions

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Herbivore pressure

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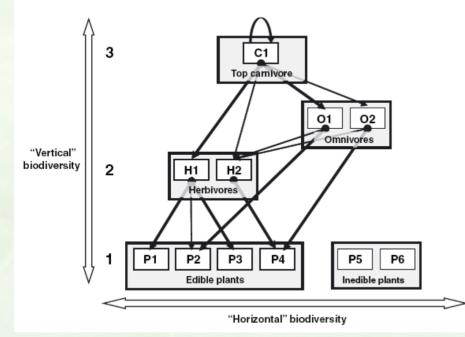
Multiple trophic level interactions

Intraguild predation, Sampling effects, Additivity & Synergisms

Duffy et al., Ecology Letters 2007

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Intraguild predation, Sampling effects, Additivity & Synergisms

→ Different impacts on the efficiency of pest control

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Agricultural production is regulated by **bottom-up** and **top-down** processes

Crop

production

Abiotic constraints Resource limitation

Availability of nutrients / Level of N fertilizer Herbivore pressure

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Research questions

- How does the level of N fertilizer impact
 - a) herbivore abundance
 - b) the efficiency of pest control
 - c) interactions between trophic levels and between guilds of natural predators

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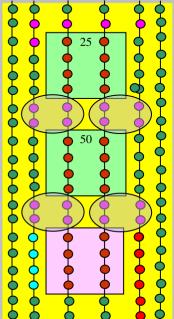
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- How does the degree of herbivore pressure and natural predator compensation impact nutrient uptake and biomass production?
- What is the relative contribution of top-down control on agricultural production, vs. bottom-up regulation?



Methods

No predators	Parasitoids	Parasitoids Ground dwellers	Birds Parasitoids	Birds Parasitoids Ground dwellers
	B + GD	в	GD	



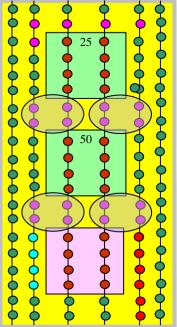


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→ Monitoring: herbivory + abundance herbivores + predators + weed cover



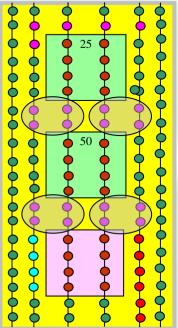




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- → Monitoring: herbivory + abundance herbivores + predators + weed cover
- → Harvest 75 days: total fresh biomass + crop fresh biomass + sellable crop fresh biomass + final weed cover + total weed biomass + C/N protocol

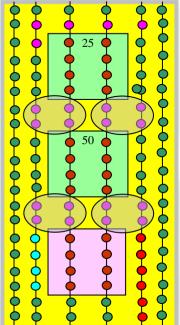




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- → Monitoring: herbivory + abundance herbivores + predators + weed cover
- → Harvest 75 days: total fresh biomass + crop fresh biomass + sellable crop fresh biomass + final weed cover + total weed biomass + C/N protocol
- → Comparison of temperature / humidity between treatments (avoid microclimatic effects)





On behalf of all the students...



THANK YOU!